**Codling Moth (Cydia pomonella)**

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**Do You Know?**

- Codling moth is the major pest of apple and pear in Utah.
- Damaging stage: larva tunnels into fruit
- Monitoring stage: adult moth
- Use of pheromone traps and the degree-day model (based on daily temperatures) are critical for determining optimal treatment timings.
- Insecticides and pheromone-based mating disruption are currently the main management tactics.
- Insecticides are targeted at newly hatched larvae and/or eggs.
- Mating disruption devices need to be applied immediately after biofix (first moth activity) to prevent or adequately delay moth mating.
- Biological control is minimally effective because larvae are protected inside fruit.
- Insect development and spray timing information are available on the USU Extension Integrated Pest Management (IPM) Pest Advisories Web page (http://utahpests.usu.edu/ipm/htm/advisories) or from your county USU Extension office.

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**Hosts**

apple, apricot, cherry, crabapple, English walnut, hawthorn, quince, pear

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Codling moth (Cydia pomonella – Family Tortricidae) is the most serious pest of apple and pear worldwide. In most commercial fruit producing regions and home yards in Utah, fruit must be protected to harvest a crop. Insecticides are a main control tactic. There are new insecticide compounds available, many of which are less toxic to humans and beneficial insects and mites than earlier insecticides. For commercial orchards with more than 10 acres of contiguous apple and pear plantings, pheromone-based mating disruption can greatly reduce codling moth populations to allow reduced insecticide use. Effective biological control has not been possible because fruit is attacked by newly hatched larvae, which are protected from natural enemies once inside the fruit. Sanitation methods can help reduce codling moth densities within an orchard but alone cannot provide satisfactory control.

In Utah, there are two to three generations of codling moth each year (Fig. 3). In northern Utah, there are typically two full generations and a partial third generation. In southern Utah, most or all of a third generation will occur. First generation moths begin to emerge about bloom time and peak in June in northern Utah. Second generation moths begin emerging in late June to early July and peak in late July to early August. Third generation moths are active from about mid August to mid September before declining day length induces the end of activity for the year.

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**Fig. 1.** Codling moth adult

**Fig. 2.** Codling moth larva

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**LIFE HISTORY**

**Pupa**
- **Size and Color:** 1/2 inch long, brown
- **Where:** pupate inside silken cocoons under bark and in other protected sites
- **7 - 30-day development period depending on temperature**

**Adult – Monitoring Stage**
- **Size:** 1/2 inch long
- **Color:** mottled gray and brown moth; bands of alternating gray and white on wings with a bronze to copper spot on the tip of each forewing (Fig. 1)
- **Where:** camouflaged against tree bark during the day with wings held tent-like over body
- **Become active when evening temperatures exceed 55 - 60° F.**
- **After emerging, male moths can be caught in traps baited with sex pheromone (codlemone) and both sexes can be caught in traps baited with codlemone and pear ester**
- **Peak moth activity is a few hours before and after twilight**
- **Mated female moths can lay 30 - 70 eggs**

**Egg**
- **Size and Shape:** pinhead sized, flat, oval
- **Color:** translucent when first deposited, later turning white; just before hatching, the black head of the larva is visible
- **Where:** laid singly on fruit or on upper surface of leaves near fruit; difficult to spot in the orchard
- **Hatch occurs in 6 - 20 days depending on temperature**

**Larva – Overwintering Stage**
- **Size and Color:** 1/2 - 3/4 inch long when full grown, creamy-white to tan
- **Where:** in silken cocoons under loose bark on tree, in protected areas at base of tree, in fruit bins, or in orchard trash piles
- **Develop to pupal stage when spring temperatures exceed 50° F in late February and early March**

**Larva – Damaging Stage**
- **Size and Color:** 1/10 inch long upon hatching, pale white with a black head; 1/2 - 3/4 inch long when full grown, tan to pink with a brown head (Fig. 2)
- **Where:** occasionally feed on terminal leaves and bore into shoots before seeking fruit
- **Bore into fruit within 24 hours after hatching, then tunnel to core where they feed on developing seeds**
- **Pass through five instars inside fruit in 3 to 5 weeks**
- **Fruit attacked early in the spring often drops; larval development can be completed in the fallen fruit**
- **After completing development, larvae exit from fruit by entry hole or by a new exit hole and crawl to a protected site for pupation**
- **Entrance and exit holes are filled with frass (excrement) and are usually conspicuous (Fig. 4)**
- **Larvae may pupate and emerge as second- or third-generation adults in 10 - 20 days, or enter diapause and remain larvae until the following spring (Fig. 3)**

**Figure 3.** Life history of codling moth. In Utah, there are three generations per year.

**HOST INJURY**

**Deep Entries**
- Larvae tunnel to center of fruit to feed on seeds (Fig. 2)
- Brown frass (excrement) extrudes from entry and exit holes (Fig. 4)
- Fruit attacked during the first generation often drops prematurely

**Stings**
- Stings are healed shallow or aborted entries that occur due to larvae death or when larvae exit the feeding area and tunnel into fruit elsewhere
Monitoring with Pheromone Traps

**Trap Placement**

- Delta or wing style pheromone traps can be used to monitor adult activity (Fig. 5).

**Sex pheromone lures** are used in traps to attract moths. There is a choice of lures available, in a rubber septum or membrane:

<table>
<thead>
<tr>
<th>Lure Type</th>
<th>Sex Attracted</th>
<th>Orchard Type</th>
<th>Lure Longevity</th>
</tr>
</thead>
<tbody>
<tr>
<td>1X (codlemone)</td>
<td>Males</td>
<td>Non-mating disrupted (MD)</td>
<td>30 or 60-day</td>
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<tr>
<td>10X (codlemone)</td>
<td>Males</td>
<td>MD</td>
<td>30-day</td>
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<tr>
<td>CM-DA combo (codlemone + pear ester)</td>
<td>Both</td>
<td>Both; primarily MD</td>
<td>60-day</td>
</tr>
</tbody>
</table>

- Place traps in orchards by first bloom or based on degree-day (temperature) accumulations (Table 1).
- Place traps within the upper third of the tree canopy (preferably 6 - 7 ft. high) making sure the trap entrance is not blocked and that it is parallel to the prevailing wind direction (Fig. 6).
- A minimum of two traps should be placed in each orchard. For orchards greater than 10 acres, place one trap for every five acres.
- Hang at least one trap on the edge and at least one near the center of the orchard to determine if moths are immigrating from outside sources and/or over wintering within the orchard. Suspected “hot spots” within the orchard should be monitored separately.
- Check traps every 1 - 2 days until the first moth is caught.

**Biofix**

- Biofix is a biological marking point from which the rest of an insect’s development is measured. It is the beginning of consistent moth flight, or where at least two moths are trapped on consecutive nights.
- It is imperative to determine the date on which biofix occurs to accurately initiate the codling moth model.

**Trap Servicing**

- Trap catch data can be used to monitor moth emergence to start degree-day accumulations, to assist with determining optimal spray timings, to determine the relative size of the moth population, and to help in evaluating the success of your management program.
- Check traps weekly and record the number of moths caught (see Codling Moth Sampling Form, ENT-13SF-06). After recording, remove moths from trap.
- Change pheromone caps based on manufacturer’s recommended product longevity and change sticky trap panels after catching 20 - 30 moths or after debris has collected on the surface.
- Zero trap catch does not necessarily mean there are no moths in the orchard. Evening temperatures below 60° F are not conducive to moth flight, and a lack of wind in the evening means the trap cannot create a pheromone plume, which lures moths inside (Fig. 6). Also, old or ineffective lures can cause zero trap catch.
- Do not cross-contaminate lures or traps between insect species. Do not handle or store unsealed pheromone lures together from more than one species. Do not reuse a trap that contained a pheromone lure from another species.
- Plan to use the same type of trap and lure from year to year so that you can compare results.
Degree-day Model

The Degree-day Method

• The development of codling moth, like all insects, can be predicted based on accumulated heat over time, called degree days (DD). Use of the codling moth phenology model based on DD will help to more accurately time insecticide applications and reduce the number of applications to a minimum.

• Codling moth development occurs between the lower and upper temperature thresholds of 50° F and 88° F.

• Starting March 1 in northern Utah or January 1 in southern Utah, begin accumulating DD for an individual location by:
  - collecting representative daily maximum and minimum air temperatures and using the DD look-up table (Table 2), or
  - obtaining the information provided by USU Extension on the IPM Pest Advisories Web page (http://utahpests.usu.edu/ipm/htm/advisories) or from your county extension office.

• Place pheromone traps in orchards when 100 DD have accumulated. The first moths are expected by 150 - 200 DD.

• Once biofix (first consistent moth catch) has occurred, accumulated DD are reset to zero (Table 1).

Timing Sprays

• If mating disruption (MD, see page 6) is used in an orchard, dispensers should be hung immediately after biofix to prevent mating and egg-laying. Supplemental insecticide treatments are usually necessary even when MD is used. The first cover spray is often the most important to apply as this timing should suppress the first generation and thus the following generations.

• Depending on the type of insecticide used, the first cover spray should be applied as follows:

<table>
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<tr>
<th>DD after biofix</th>
<th>Timing/Target</th>
<th>Examples</th>
</tr>
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<tbody>
<tr>
<td>50 - 75</td>
<td>pre-egg-laying</td>
<td>Rimon</td>
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<tr>
<td>100 - 200</td>
<td>early egg-laying</td>
<td>Horticultural oil, Esteem, Intrepid</td>
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<tr>
<td>220 - 250</td>
<td>first egg hatch (emergence of larvae)</td>
<td>Assail, Asana, Calypso, Carbaryl, Diazanon, Guthion, Codling Moth Granulosis Virus, Imidan</td>
</tr>
</tbody>
</table>

• Reapply insecticides based on the residual period (i.e., protection interval) of the product used. Keep fruit protected throughout each generation (Table 1).

• As harvest date approaches, consider the pre-harvest interval (required time interval between insecticide application and harvest) in planning late season treatments.

Table 1. Major events in a codling moth management program, based on accumulated degree days

<table>
<thead>
<tr>
<th>Degree Days</th>
<th>% Adults</th>
<th>% Eggs</th>
<th>Management Event</th>
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<tbody>
<tr>
<td>100 *</td>
<td>0</td>
<td>0</td>
<td>Place traps in orchards</td>
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<tr>
<td>150 - 200</td>
<td>First moths expected</td>
<td>0</td>
<td>Check traps every 1-2 days until biofix is determined</td>
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</tbody>
</table>

First Generation

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<tr>
<th>(biofix)</th>
<th>First consistent catch</th>
<th>0</th>
<th>Reset degree days to 0</th>
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<tbody>
<tr>
<td>50 - 75</td>
<td>5 - 9</td>
<td>0</td>
<td>First eggs are laid</td>
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<tr>
<td>100 - 200</td>
<td>15 - 40</td>
<td>0</td>
<td>Apply insecticides that need to be present before egg-laying</td>
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<tr>
<td>220 - 250</td>
<td>45 - 50</td>
<td>1 - 3</td>
<td>Early egg-laying period</td>
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<td>340 - 640</td>
<td>67 - 98</td>
<td>12 - 80</td>
<td>Apply insecticides that target newly hatched larvae</td>
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<tr>
<td>920</td>
<td>100</td>
<td>99</td>
<td>End of egg hatch for 1st generation</td>
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</tbody>
</table>

Second Generation

| 1000 - 1050 | 5 - 8 | 0 | First eggs of 2nd generation are laid |
| 1100         | 13    | 1 | Apply insecticides to target early egg-laying |
| 1320 - 1720  | 46 - 93 | 11 - 71 | Critical period for control, high rate of egg hatch |
| 2100         | 100   | 99 | End of egg hatch for 2nd generation |

Third Generation

| 2160 | 1 | 15 | Beginning of egg hatch |
|      |   |   | Keep fruit protected through September 15 |
|      |   |   | Check pre-harvest interval of material used to ensure that final spray is not too near harvest |

*Begin accumulating degree days after daily temperatures begin to exceed 50°F, typically on January 1 for southern Utah or March 1 for northern Utah.

† Biofix is when at least 2 moths are caught on consecutive nights.
Insecticides

Synthetic insecticides have been the major control tactic used since the 1940s. Current insecticide choices include synthetic materials, microbial and botanical insecticides, and petroleum oils. The choice depends on numerous factors including commercial versus home orchard production, the crop’s market destination, grower preferences, size of orchard, codling moth pressure in the area, and the surrounding habitat. Select insecticides with the desired modes of action and apply them at optimal timings to coincide with key development periods (Table 1). Use of pheromone trapping in combination with the degree-day model is highly recommended to accurately determine codling moth development for your location. (For more information, see the Degree-day Method above.)

Table 2. Degree Day Look-Up for Codling Moth*

Lower threshold: 50° F  Upper threshold: 88° F

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<thead>
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<th>Minimum Temperature</th>
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To find the total degree days for a day, locate the low and high temperatures and follow the rows to where they intersect. For temperatures between those listed, use the nearest shown.


MANAGEMENT
If the orchard is large (more than 10 acres) and moths are only found in border traps, treating 4 - 5 border rows may markedly reduce the amount and number of insecticide applications. This tactic is primarily used after first and second cover sprays have been applied or in conjunction with mating disruption.

Rotate among insecticide classes (see list of insecticides below) to prevent development of resistance to insecticides in the codling moth population.

**Synthetic Insecticides***

Chloronicotinyls:
- acetamiprid (Assail)
- clothianidin (Clutch)
- thiacloprid (Calypso)

Insect Growth Regulators:
- azadirachtin (Aza-Direct, Azatin)**
- methoxyfenozide (Intrepid)
- novaluron (Rimon)
- pyriproxyfen (Esteem)
- tebufenozide (Confirm)

Organophosphates:
- azinphosmethyl (Guthion)
- chlorpyrifos (Lorsban), apples only
- diazinon (Diazinon)
- dimethoate (Dimethoate)
- malathion (Malathion)**
- phosmet (Imidan)

Carbamates:
- carbaryl (Sevin)**
- indoxacarb (Avaunt)

Synthetic pyrethroids:
- esfenvalerate (Asana)
- fenpropathrin (Danitol)
- lambda-cyhalothrin (Warrior)

**Lower Toxicity Insecticides***

Use of lower toxicity insecticides alone has not generally provided satisfactory control of codling moth. However, intensive use of combinations of soft pesticides has proven adequate in some cases. Combinations of soft chemicals and pheromone-based mating disruption have proven effective.

**Horticultural mineral oils***:
Highly refined, superior-type petroleum oils can prevent egg hatch by suffocation. Apply at beginning of early egg-laying (100 - 200 and 1000 - 1050 DD after biofix). There are concerns about negative effects on fruit finish following use of high rates and multiple applications.

**Microbial insecticides**:
- *Bacillus thuringiensis* (Dipel**, Javelin, Biobit, Crymax), codling moth granulosis virus (Virossoft, Cyd-X, Carpo-

**Particle films**:
Kaolin clay (Surround**), when maintained as a complete barrier on fruit surfaces, can reduce codling moth infestations.

**Botanical insecticides**:
- rotenone**, pyrethrum**, and ryania** (apples only) have variable control effectiveness. They may provide satisfactory control of low codling moth population levels but can be harsh on beneficials, thus allowing other pest insect populations to increase.

*All brand names are registered trademarks. Examples of brands may not be all-inclusive, but are meant to provide examples of effective insecticides registered on apple and pear in Utah. The availability of insecticides is changing rapidly. Always check the label for registered uses, application and safety information, and protection and pre-harvest intervals.

**Insecticide products that may also be available for use on home fruit trees.

**Mating Disruption**

Detailed knowledge of insect biology, development timings, and limitations of mating disruption are needed to use it successfully. Substantial fruit damage could result from improper use. The size of orchard, proximity to outside sources of codling moth, and dispenser placement and application rate can all influence the success of this tactic. The typically small size of apple and pear blocks in Utah (less than 10 acres) and the high codling moth populations reduce the effectiveness of mating disruption in Utah as compared to other areas of the Northwest. Mating disruption is not effective in home yards or orchards smaller than 10 acres.

- Place small dispensers containing the female sex pheromone (Isomate C+, Isomate CTT, No Mate, CheckMate) at 200 - 400 dispensers per acre (follow product label directions) throughout the orchard immediately after biofix. Place dispensers within 2 feet of the top of the canopy. Sprayable formulations and aerosol devices (also called puffers) for releasing pheromones are available, but experience with these products is limited.

- Research in Utah and other locations has demonstrated the importance of continual monitoring of adult populations with traps in mating disrupted (MD) orchards. The dual gender lure (CM-DA Combo®, Pherocon) has proven more effective than the 10X codlemone lure in catching moths in MD orchards.

- If the orchard has a history of codling moth problems, use one or two applications of insecticides against the first generation. If a codling moth source exists nearby, use border sprays (5 - 6 rows) of insecticides to prevent fruit damage on edges from immigrating mated moths.
• Be aware that if insecticide applications are reduced substantially, populations of other pests (e.g., lea-frollers, piercing-sucking bugs, aphids, scale, etc.) can increase.

**Fruit Thinning**
Newly hatched larvae often seek out protected sites for entry, and thinning fruit to just one apple/cluster can limit successful entries. Fruit thinning also allows for improved insecticide coverage on the entire fruits.

**Sanitation**
- Remove or treat host trees within a quarter mile (450 yards) of orchards to destroy outside codling moth sources, including abandoned orchards and wild hosts.
- Strip fruit remaining after harvest in young, unharvested orchards or on pollinator trees.
- Remove or destroy piles of culled fruit in orchards.
- Remove additional pupation sites from orchards such as fruit bins, brush, woodpiles, and other debris.
- Fruit infested during the first generation typically drop to the ground in June or July. Remove or destroy (e.g., flail) dropped fruit to reduce second generation densities.

**Trunk Banding**
- Place corrugated cardboard bands (2 - 3 inches wide) around trunks of trees in May to collect first generation larvae or in August to collect overwintering larvae that are moving to the trunks to pupate.
- Remove and destroy bands before moths emerge in mid- to late June (for first generation) or in late October to November (for overwintering generation).
- This method is most effective on smooth-barked varieties and in smaller, isolated orchards without nearby sources of codling moth.

**Biological Control**
Use of more selective and lower toxicity insecticides enhances populations of beneficials (predaceous and parasitic arthropods). Native enemies alone, however, do not provide satisfactory control. Recent efforts to introduce parasitoids from native habitats of codling moth in Eurasia are promising. The egg parasitoid, *Trichogramma*, has shown potential especially in combination with other lower toxicity tactics.

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Precautionary Statement: All pesticides have benefits and risks, however following the label will maximize the benefits and reduce risks. Pay attention to the directions for use and follow precautionary statements. Pesticide labels are considered legal documents containing instructions and limitations. Inconsistent use of the product or disregarding the label is a violation of both federal and state laws. The pesticide applicator is legally responsible for proper use.

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